



AssayMax Human Factor IX (FIX) ELISA Kit

Catalog Number EF1009-1

Introduction

Factor IX (FIX) is zymogens of plasma serine proteases required for normal hemostasis (1). FIX and FX were activated by tissue factor (TF) and factor VIIa (FVIIa) complexes and initiates coagulation resulting in thrombin formation (2). Hemophilia B is an X-linked bleeding disorder that results from a deficiency in functional coagulation factor IX (hFIX) (3). On the other hand, increased plasma level of FIX was reported to be independent risk factor of venous thromboembolism (VTE) (4).

Principal of the Assay

The AssayMax Human Factor IX (FIX) ELISA kit is designed for detection of human factor IX in plasma and cell culture supernatants. This assay employs a quantitative sandwich enzyme immunoassay technique that measures FIX in 3.5 hours. A polyclonal antibody specific for FIX has been pre-coated onto a 96-well microplate with removable strips. FIX in standards and samples is sandwiched by the immobilized antibody and the biotinylated polyclonal antibody specific for FIX, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- This kit is for research use only.
- The kit should not be used beyond the expiration date.
- The Stop Solution is an acid solution.

Reagents

- **FIX Microplate:** A 96 well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against FIX.
- **Sealing Tapes:** Each kit contains 3 pre-cut, pressure-sensitive sealing tapes that can be cut to fit the format of the individual assay.
- **FIX Standard:** Human FIX in a buffered protein base (800 ng, lyophilized).
- **Biotinylated FIX Antibody (100x):** A 100-fold concentrated biotinylated polyclonal antibody against FIX (80 µl).
- **EIA Diluent Concentrate (10x):** A 10-fold concentrated buffered protein base (30 ml).
- **Wash Buffer Concentrate (20x):** A 20-fold concentrated buffered surfactant (30 ml).
- **Streptavidin-Peroxidase Conjugate (SP Conjugate):** A 100-fold concentrate (90 µl).

- **Chromogen Substrate:** A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- **Stop Solution:** A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Store unopened kit at 2 - 8⁰C up to expiration date.
- Opened EIA Diluent may be stored for up to 1 month at 2-8⁰C. Store reconstituted reagents at -20⁰C or below.
- Opened unused strip wells may return to the foil pouch with the desiccant pack, reseal along zip-seal. May be stored for up to 1 month in a vacuum desiccator.

Other Supplies required

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 µl, 20-200 µl, 200-1000 µl and multiple channel pipettes).
- Deionized or distilled reagent grade water.

Sample Collection, Preparation and Storage

- **Plasma:** Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 2000 x g for 10 minutes and collect supernatants. Dilute samples 1:400 into EIA Diluent and assay. The undiluted samples can be stored at -20⁰C or below for up to 3 months. Avoid repeated freeze-thaw cycles. (EDTA or Heparin can also be used as anticoagulant.)
- **Cell Culture Supernatants:** Collect cell culture media and centrifuge at 2000 x g for 10 minutes at 4⁰C to remove debris. Collect supernatants and assay. The undiluted samples can be stored at -20⁰C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use.
- **EIA Diluent Concentrate (10x):** Dilute the EIA Diluent 1:10 with reagent grade water. Store for up to 1 month at 2-8⁰C.
- **FIX Standard:** Reconstitute the 800 ng of human FIX Standard with 4 ml of EIA Diluent to generate a standard solution of 200 ng/ml. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare triplicate standard points by serially diluting the Standard solution (200 ng/ml) 1:2 with EIA Diluent to produce 100, 50, 25, 12.5, 6.25, and 3.13 ng/ml. EIA Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20⁰C.

Standard Point	Dilution	[FIX] (ng/ml)
P1	1 part Standard (200 ng/ml)	200.00
P2	1 part P1 + 1 part EIA Diluent	100.00
P3	1 part P2 + 1 part EIA Diluent	50.00
P4	1 part P3 + 1 part EIA Diluent	25.00
P5	1 part P4 + 1 part EIA Diluent	12.50
P6	1 part P5 + 1 part EIA Diluent	6.25
P7	1 part P6 + 1 part EIA Diluent	3.13
P8	EIA Diluent	0.00

- **Biotinylated FIX Antibody (100x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:100 with EIA Diluent. Any remaining solution should be frozen at -20°C.
- **Wash Buffer Concentrate (20x):** Dilute the Wash Buffer Concentrate 1:20 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with EIA Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

- Prepare all reagents, working standards and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-30°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccant inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 50 µl of Standard or sample per well. Cover wells with a sealing tape and incubate for two hours. Start the timer after the last sample addition.
- Wash five times with 200 µl of Wash Buffer. Invert the plate and decant the contents, and hit the plate 4-5 times on absorbent paper towel to complete remove liquid at each step.
- Add 50 µl of Biotinylated FIX Antibody to each well and incubate for one hour.
- Wash five times with 200 µl of Wash Buffer as above.
- Add 50 µl of Streptavidin-Peroxidase Conjugate per well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash five times with 200 µl of Wash Buffer as above.
- Add 50 µl of Chromogen Substrate per well and incubate for about 10 minutes or till the optimal color density develops. Gently tap the plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 µl of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm **immediately**. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

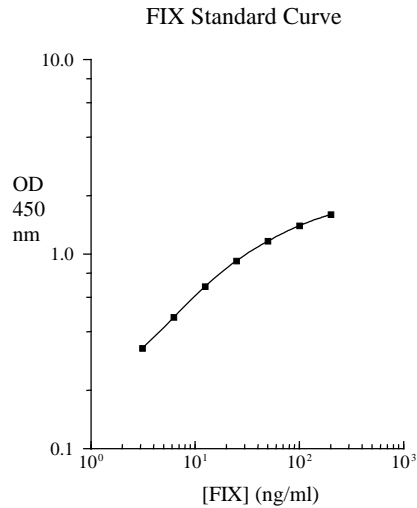
Data Analysis

- Calculate the mean value of the triplicate for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.

- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

Standard Curve

- The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



Performance Characteristics

- The minimum detectable level of FIX was typically less than 2 ng/ml.
- Intra-assay and inter-assay coefficients of variation were 5.8% and 6.9% respectively.

Linearity

	Average Percentage of Expected Value
Sample Dilution	Plasma
1:200	97%
1:400	100%
1:800	102%

Recovery

Standard Added Value	5 – 100 ng/ml
Recovery %	88-110 %
Average Recovery %	99 %

Cross-Reactivity

Species	% Cross Reactivity
Beagle	None
Bovine	None
Monkey	< 10 (suggest dilution 1:4 for plasma)
Mouse	None
Rat	None
Swine	None

- 10% FBS in culture media will not affect the assay.

References

- (1) Wang X. *et al.* (2005) *J Thromb Haemost.* 3(4): 695-702
- (2) Bogdanov VY *et al* (2003) *Nat Med.* 9(4): 458-62
- (3) Pipe SW (2004) *Semin Thromb Hemost.* 30(2): 227-37
- (4) Bertina RM. (2003) *Pathophysiol Haemost Thromb.* 33(5-6): 395-400.

Version 4.2